

# How to access and efficiently perform crystallographic fragment screening at XChem

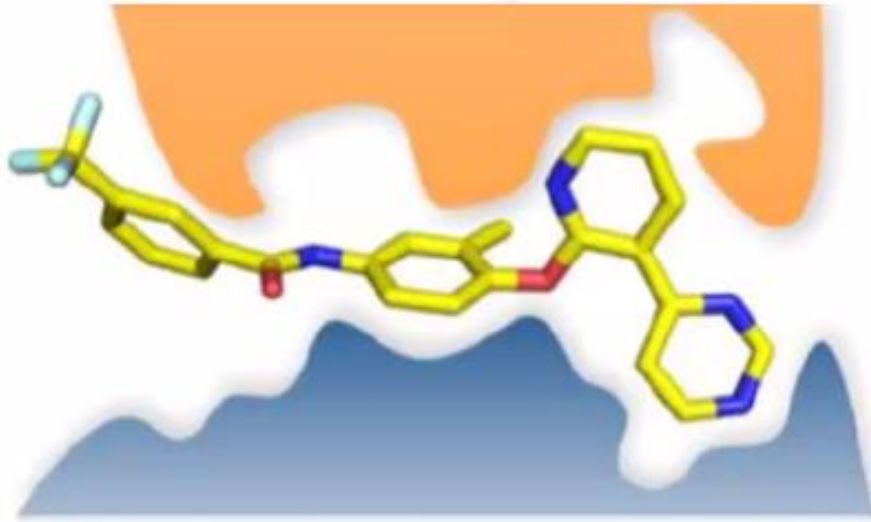


Halina Mikolajek

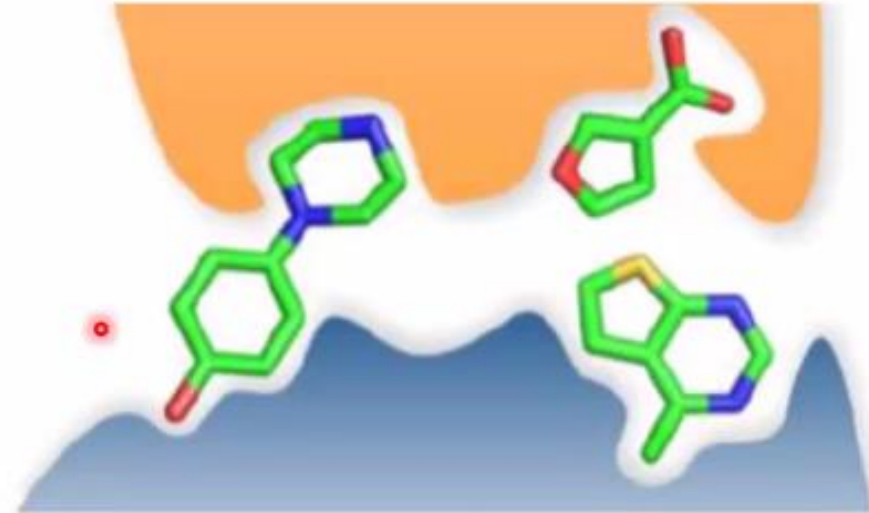


Daren Fearon

# HTS vs Fragment-Based Drug Discovery

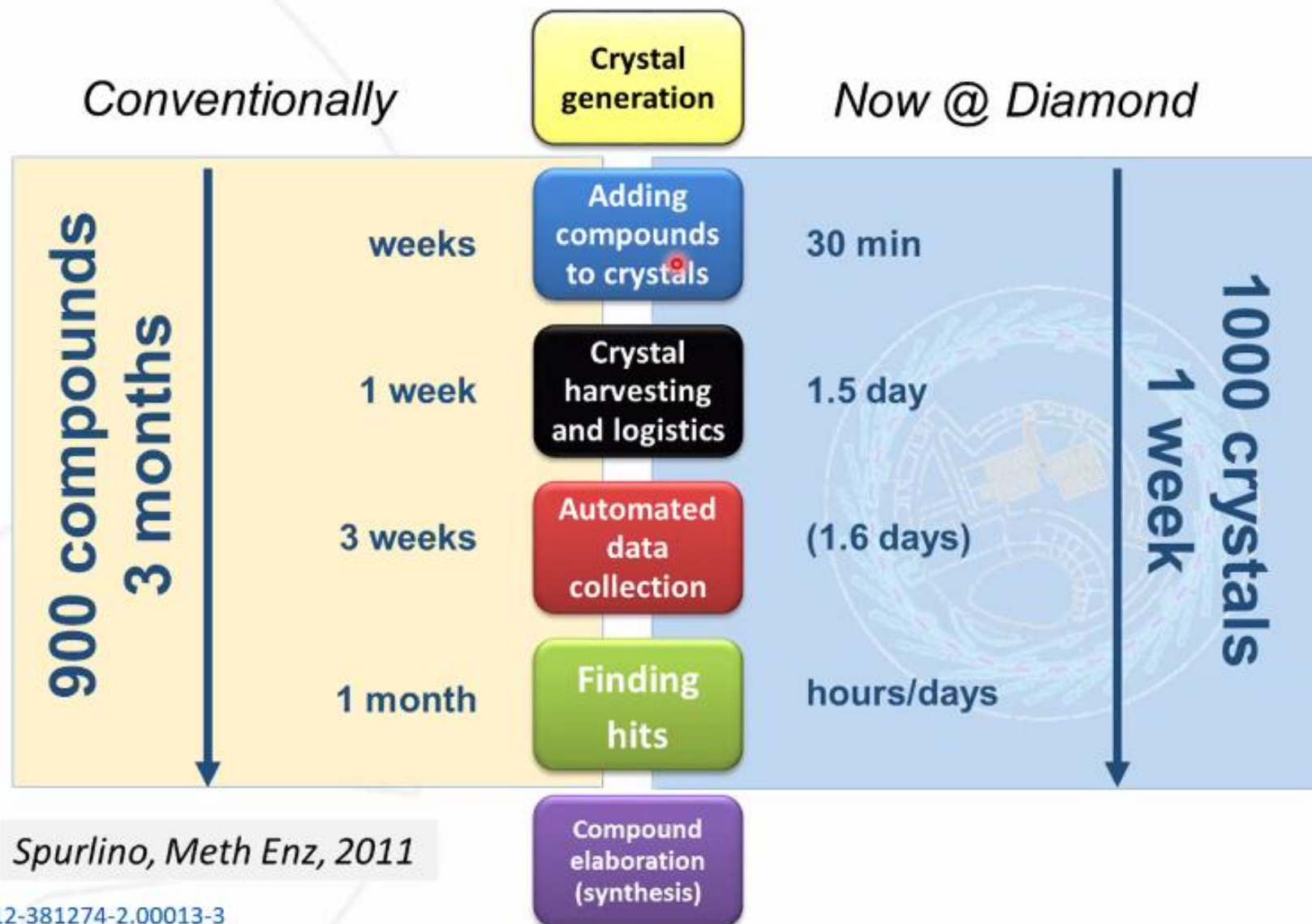


- Libraries typically >100,000 compounds
- Molecular weight >300 Da
- Affinities typically in  $\mu\text{M}$ -nM range
- Usually identified by biochemical or cellular assays
- Can be difficult to optimise



- Libraries typically 500-1000 compounds
- Molecular weight < 250 Da
- Affinities typically in mM- $\mu\text{M}$  range **BUT** compounds are very efficient
- Usually identified by biophysical methods
- Iterative optimisation driven by structure-based methods

# Fragment screening at Diamond: XChem



# XChem Workflow



Crystal imaging  
and ranking



Compound  
dispensing



Sample  
harvesting



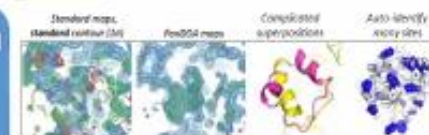
Unattended  
data collection



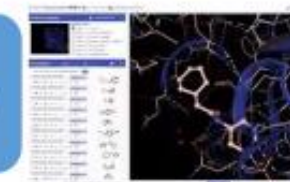
Automated data  
processing



Hit identification  
(PanDDA) and  
modelling



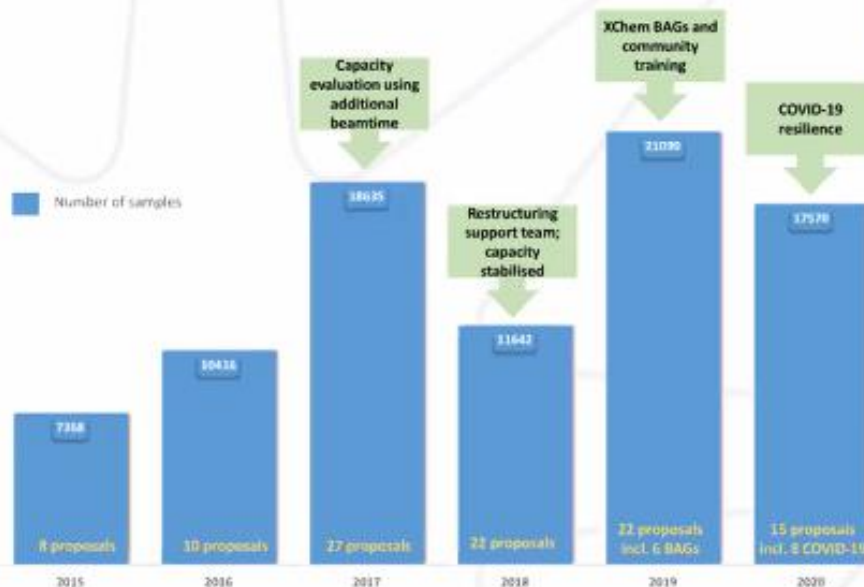
Model  
dissemination  
(Fragalysis)



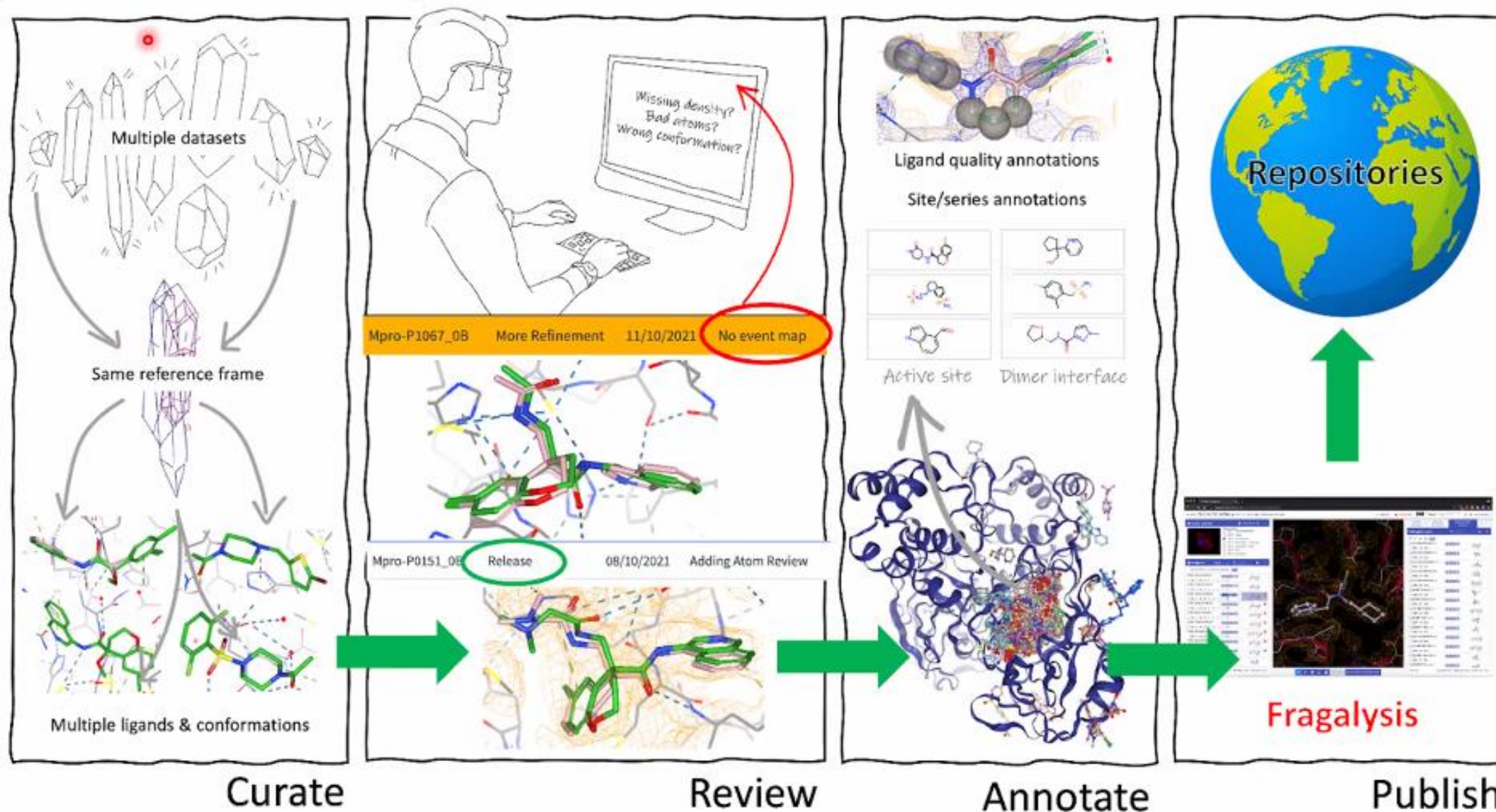
Well curated libraries  
Best practice protocols

Streamlining in MX village  
Technology for user labs

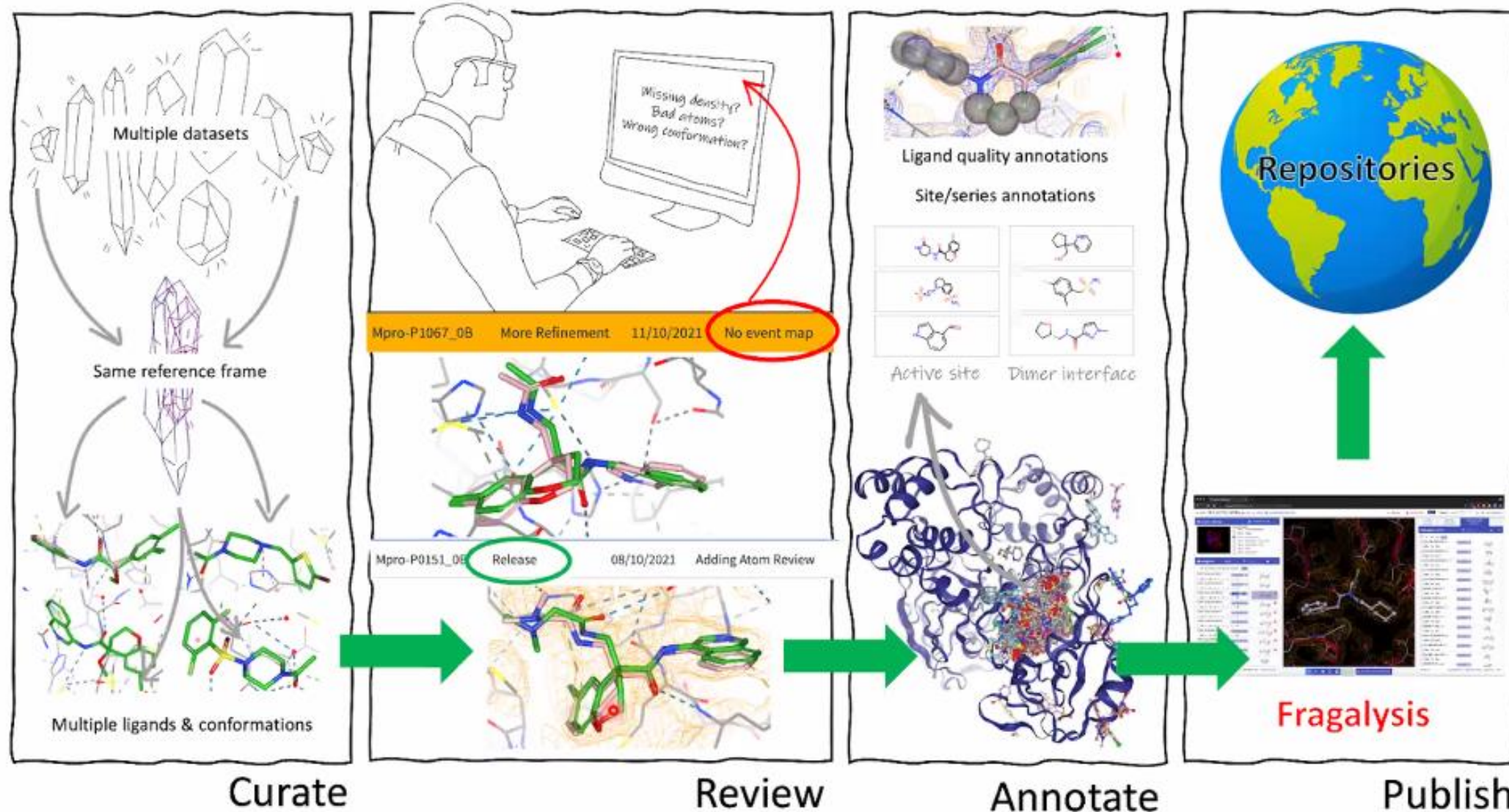
New algorithms  
New platforms



# Data Review & Dissemination

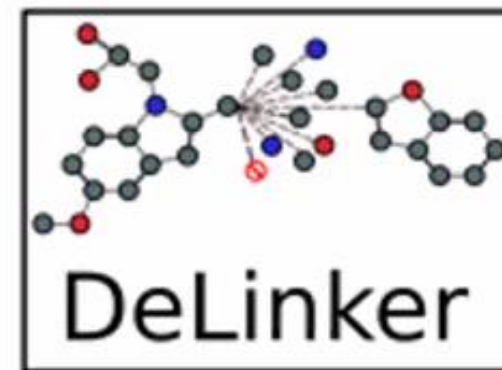
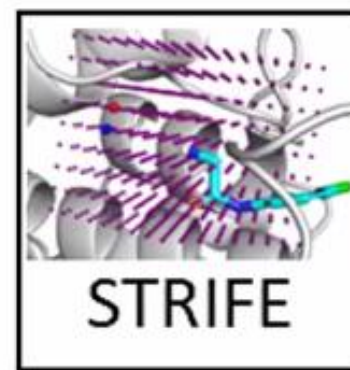
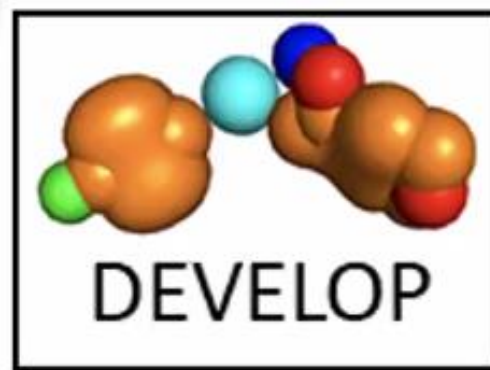
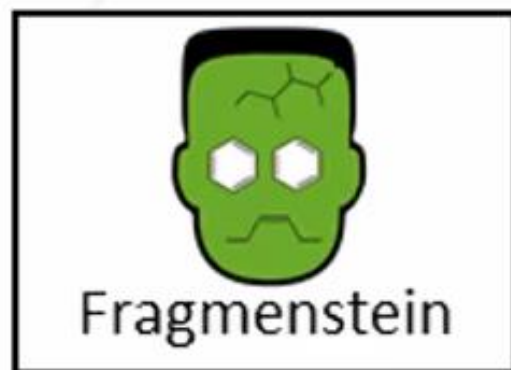
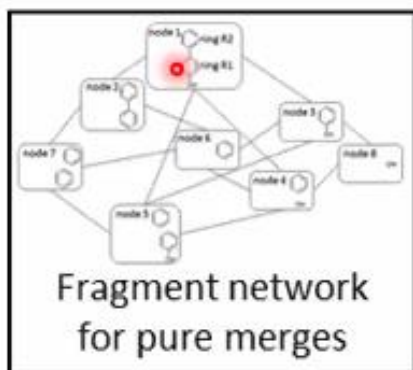
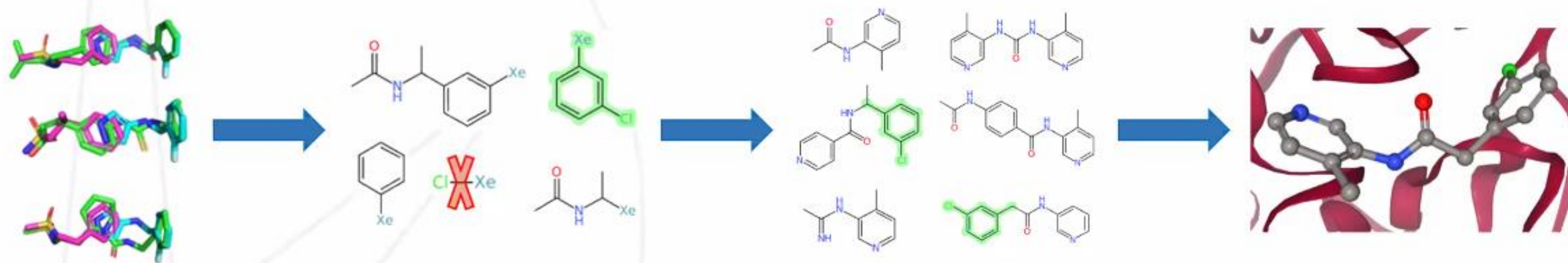


# Data Review & Dissemination



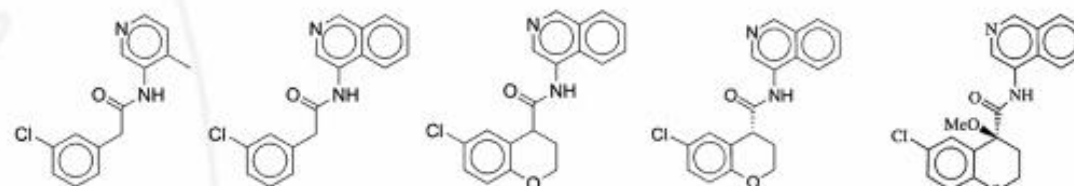
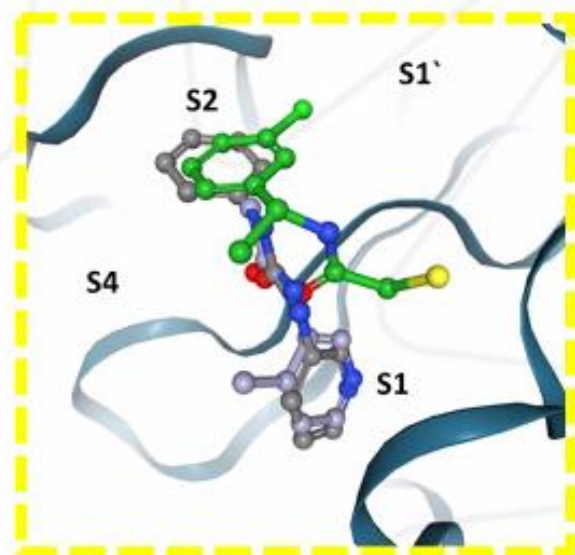
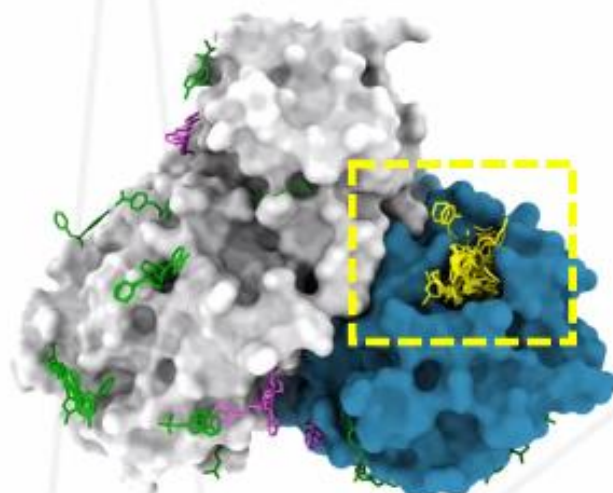
# Algorithmic Fragment Progression

- Computational tools being developed to accelerate routine hit-to-lead

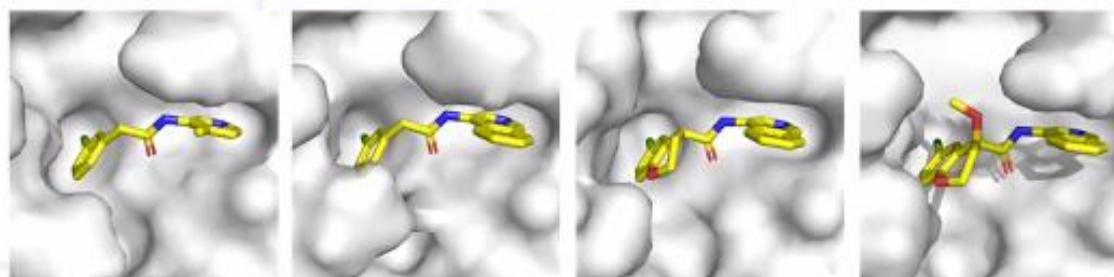


# What results can I expect?

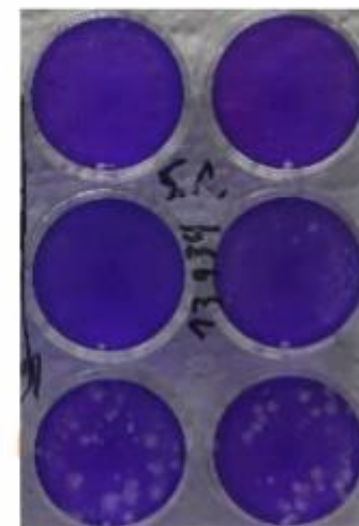
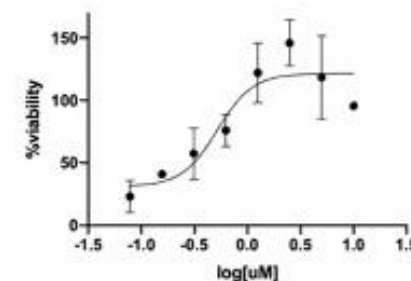
- Merging fragments can rapidly lead to potency



TRY-UNI-714a760b-6  $IC_{50}=24 \mu M$  ADA-UCB-6c2cb422-1  $IC_{50}=720 nM$  VLA-UCB-1dbca3b4-15  $IC_{50}=360 nM$  MAT-POS-b3e365b9-1  $IC_{50}=140 nM$  PET-UNK-29afea89-2  $IC_{50}=80 nM$



Lead compound active  
against live SARS-CoV-2



# How to access - Standard Academic Access



- Covers single target with calls issued twice a year (usually Apr and Oct)
  - Proposals evaluated by an independent peer-review panel
  - Includes iNEXT-Discovery applications
- Tier 1: Exploratory projects (100-200 fragments)
  - Feasibility is high (robust crystal system established) but credible strategy for follow-up of hits is not in place
- Tier 2: Full screen (700-1000 fragments)
  - Feasibility has been demonstrated and a robust strategy for progressing hits has been provided
- Tier 3 (**new**): Follow-up support (200-300 compounds)
  - Feasibility previously established and design rationale articulated for peer review

# Academic BAG Access

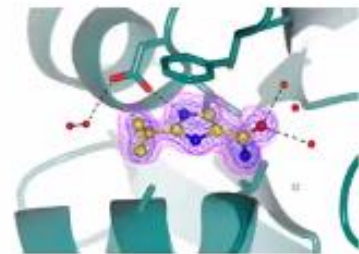
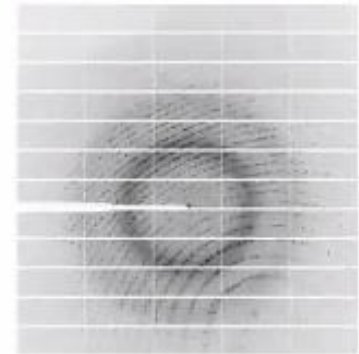
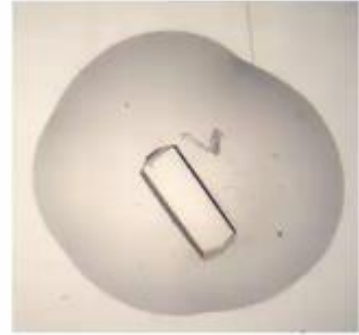
For groups, institutes or collaborations that:

- Have a pipeline of projects all aimed specifically at compound development
- Have hit-to-lead infrastructure in place
- Will therefore routinely and predictably have crystal systems for evaluation and screening
- Have a stringent internal prioritisation process
- Have experienced XChem crystallographer (super-user) to organise and provide logistical support to BAG members

New BAG applications welcome!

# Ideal XChem ready crystal systems

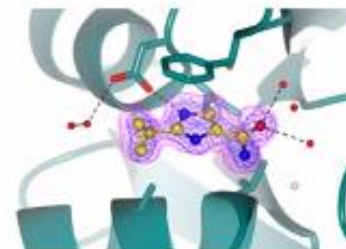
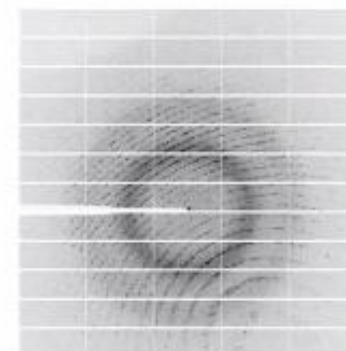
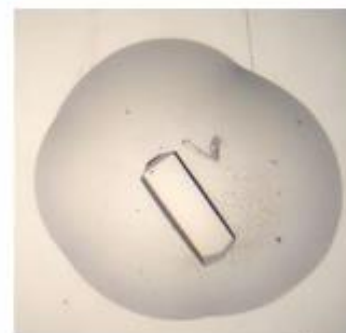
- Grow reproducibly (>50% drops) in SWISSCI 3-drop plates
- Are chunky, rather than needles
- Consistently diffract to high resolution (<2.5 Å)
- Tolerate high solvent concentrations
- Don't stick to the plate
- Don't grow skin on the drop
- Don't require complicated cryoprotection



**But non-ideal crystals are feasible!**

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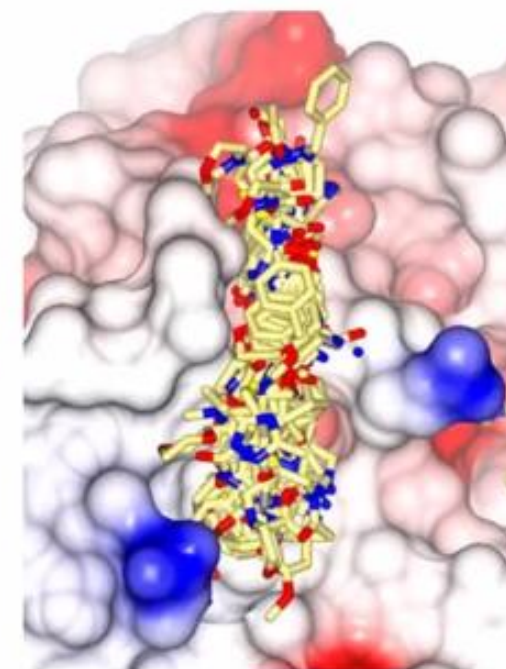
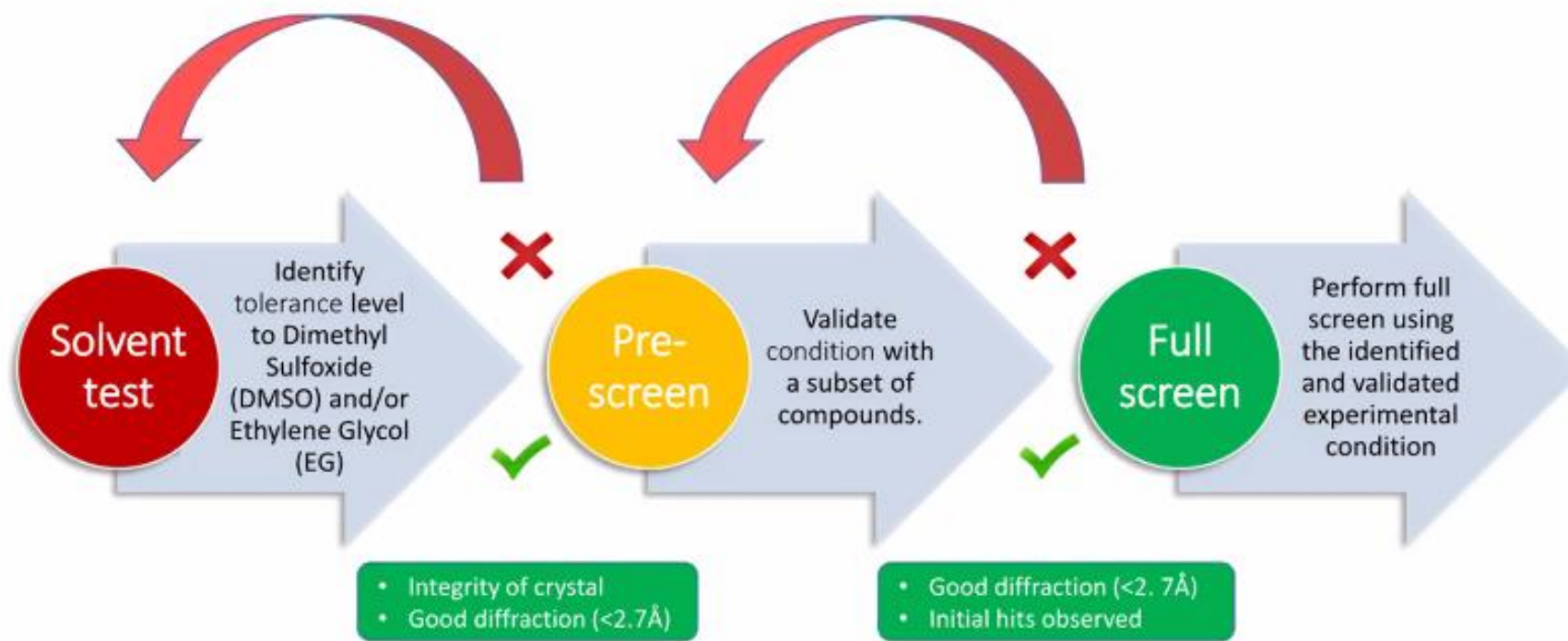


**But non-ideal crystals are feasible!**

# Establishing “XChem ready” systems

- Explore multiple protein constructs and protein engineering
- Identify multiple crystallisation conditions/crystal forms
  - PEG preferable over high salt conditions
  - Be aware of pH and volatile solvents
- Run QC for your protein batches and crystal trays
- Use seeding!
- Determine crystal solvent tolerance and life span of crystals
- Test transferability of crystallisation/trays between locations
- When robustness established, keep things consistent

# Flushing out the problems



Screening outcome

## Typical soaking experiments

- (30-50 drops of crystals)
- 5, 10, 20, 30% DMSO or ethylene glycol
  - 1h and 3h time course

## Typical soaking experiments

- (100-150 drops of crystals)
- Addition of compounds

# Partnerships (hit to lead follow-ups):



- EUBOpen
  - Collaboration with members of the consortium (including 5 pharma partners) to help progress hits to chemical probes
  - Can include screening assays, crystallisation, HTS, chemistry etc
  - The target remains confidential between the applicant and collaborator(s)
- EU-OPENSOURCE
  - Offers open access to its shared resources including 140,000 compounds linked to EU-OS Fragment library and med. chem. support



# Useful Contacts and Links

- Frank von Delft (PBS): 8997, [frank.von-delft@diamond.ac.uk](mailto:frank.von-delft@diamond.ac.uk)
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- Tyler Gorrie-Stone (Software engineer): [tyler.gorrie-stone@diamond.ac.uk](mailto:tyler.gorrie-stone@diamond.ac.uk)
- <http://www.diamond.ac.uk/Beamlines/Mx/Fragment-Screening.html>
- Achieving-efficient-fragment-screening-at-XChem-facility-at-Diamond Light  
Source: <https://dx.doi.org/10.3791/62414>